

HOW MANY SPECIES OF RATTLESNAKES ARE THERE IN THE *CROTALUS DURISSUS* SPECIES GROUP (SERPENTES: CROTALIDAE)?

¿CUÁNTAS ESPECIES DE CASCABELES HAY EN EL GRUPO DE *CROTALUS DURISSUS* (SERPENTES: CROTALIDAE)?

JACOBO REYES-VELASCO^{1,2*}, CHRISTIAN L. COX³, JASON M. JONES², MIGUEL BORJA⁴ & JONATHAN A. CAMPBELL⁵

¹Entorno Biótico A.C. – Colima, Colima, México.

²Herp.mx A.C. – Villa de Álvarez, Colima, México.

³Florida International University – Miami, Florida.

⁴Facultad de Ciencias Biológicas, Universidad Juárez del Estado de Durango.

⁵University of Texas at Arlington – Arlington, Texas.

*Correspondence: jacobo@Crotalus.com

Received: 2021-08-09. Accepted: 2021-11-02. Published: 2022-01-27.

Editor: Julian A. Velasco, México.

Resumen.— Los miembros del grupo de especies de *Crotalus durissus* se encuentran entre las especies más grandes de serpientes de cascabel y tienen una gran importancia médica. La taxonomía del grupo es compleja, y la línea de lo que se considera especie, subespecie o población es difícil de definir. Un estudio reciente dividió a uno de los miembros del grupo (*Crotalus culminatus*) en tres especies, la especie nominal, así como *C. ehecatl* y *C. mictlantecuhtli*, basándose en datos genéticos y morfológicos. En este estudio reanalizamos los datos mitocondriales y nucleares publicados anteriormente, así como secuencias adicionales de dos genes mitocondriales de miembros del grupo de *Crotalus durissus*, para probar si el aislamiento por distancia (y no especiación) podría ser el responsable de la variación genética observada en estudios anteriores. Nuestros resultados muestran que las diferencias genéticas entre algunas de las especies de este grupo (*Crotalus simus* – *C. durissus* y *C. culminatus* – *C. ehecatl*) son menores que las distancias genéticas intraespecíficas. El único gen nuclear disponible para el grupo *Crotalus durissus* (factor de maduración de ovocitos Mos, c-mos) no es filogenéticamente informativo y no puede distinguir entre muchas de las especies del género, por lo que es de poca utilidad a la hora de tomar decisiones taxonómicas. Encontramos una baja divergencia a nivel mitocondrial entre estos dos pares de especies, y al menos en uno de los pares de especies encontramos un patrón de aislamiento por distancia. Adicionalmente, realizamos un análisis de delimitación de especies con un método desarrollado recientemente, DELINEATE, el cual apoya la validez de *C. mictlantecuhtli*, pero no de *C. ehecatl*. Estos resultados sugieren que las diferencias moleculares a nivel mitocondrial en los miembros del grupo de especies *Crotalus durissus* podrían ser el resultado de aislamiento por distancia y no necesariamente debidos a eventos de especiación. Creemos que un mayor muestreo y datos moleculares adicionales son necesarios para aclarar los límites de las especies del grupo de *Crotalus durissus*.

Palabras clave.— ADN mitocondrial, aislamiento por distancia, límites de especies, Mesoamérica.

Abstract.— Members of the *Crotalus durissus* species group are amongst the largest species of rattlesnakes and are of strong medical importance. The taxonomy of the group is convoluted, and the line of what is considered a species, subspecies or populations is hard to define. A recent study split one of the members of the group (*C. culminatus*) into three species, the nominal species, as well as *C. ehecatl* and *C. mictlantecuhtli*, based on genetic and morphological data. Here we reanalyze previously published mitochondrial and nuclear data, as well as additional sequences for two mitochondrial genes from members of the *Crotalus durissus* group, to test

if isolation by distance (and not speciation) could be responsible for the genetic variation observed in previous studies. Our results show that the genetic differences between some of the species in the group (*Crotalus simus* – *C. durissus* and *C. culminatus* – *C. ehecatl*) are lower than intra-specific genetic distances. The only nuclear gene at hand for the *C. durissus* group (oocyte maturation factor Mos, *c-mos*) is not phylogenetically informative and cannot distinguish between many of the species in the genus, thus it is of little support when it comes to taxonomic decisions. We find low divergence at the mitochondrial level between these two species pairs, and at least in one of the species pairs we find a pattern of isolation-by-distance (IBD). Additionally, we performed a newly developed species delimitation analysis, DELINEATE, which supports the validity of *C. mictlantecuhtli*, but not *C. ehecatl*. These results suggest that the molecular differences at the mitochondrial level in members of the *Crotalus durissus* species group could be the result of IBD and not necessarily due to speciation events. We believe that additional sampling as well as additional molecular data is necessary to clarify species limits in the *Crotalus durissus* species group.

Keywords.— Isolation-by-distance, Mesoamerica, mtDNA, species limits.

INTRODUCTION

In recent years, the general lineage concept (de Queiroz, 1998) has become widely used for defining species. Under the general lineage species concept, species can be defined as separately evolving metapopulation lineages. The debate has now shifted from “what is a species?”, to “which lines of evidence are necessary to define those separately evolving lineages?” (de Queiroz, 2007). Morphological characters have traditionally been the standard when defining species, but these characters can evolve at different rates between taxa, and it can be difficult to determine what characters are diagnostic between species (Fujita et al., 2012). The use of molecular markers has become commonplace when delimiting and defining species, but the results of those studies that define species based only on molecular data alone (e.g. Card, 2016; Leaché & Fujita, 2010; Ruane et al., 2014) can be misleading when not analyzed carefully, as they might delimit genetic structure and not necessarily species (Chambers & Hillis, 2020; Sukumaran et al., 2021; Sukumaran & Knowles, 2017). Mitochondrial DNA (mtDNA) has become a standard tool in population genetics, as it can allow us to better understand inter and intra-population processes such as migration, gene flow, genetic structure, or speciation (Rubinoff & Holland, 2005). However, in recent years, the use of mitochondrial markers as the only evidence to define species has become commonplace (Hillis, 2019). A multitude of phenomena, such as rapid lineage sorting, selection, male-biased dispersal, isolation-by-distance, or introgression, could in many cases make the use of mtDNA alone unreliable for delimiting and defining species (Rubinoff & Holland, 2005).

The level of genetic variation that needs to exist between two lineages to be considered two different species is a matter of great debate, and probably there will not be a single correct answer for all cases (Sukumaran et al., 2021). However, because species are

by definition hypotheses (de Queiroz, 2007), the power to support or refute a particular hypothesis increases with the number of concurrent pieces of evidence. Species delimitation should thus be performed with an “integrative taxonomy” approach (Padial et al., 2010; Pante et al., 2015; Will et al., 2005), which means to include multiple lines of evidence when describing species, for example ecology, genetics, morphology, physiology, among others, rather than multiple analyses of the same data (Fujita et al., 2012).

Rattlesnakes of the *Crotalus durissus* group are among the largest species in the genus and their venom is of important medical concern (Campbell & Lamar, 2004). Members of this group were considered for many years a single species, *Crotalus durissus* Linnaeus, 1758, with multiple subspecies that ranged from Nuevo León and Tamaulipas in Mexico to central Argentina (Klauber, 1952; Fig. 1). The northernmost member of the group was originally described as *Crotalus totonacus* (Gloyd & Kauffeld, 1940), however Smith and Taylor (1945) considered this population a subspecies of *C. durissus* (*C. durissus totonacus*). This designation was followed by multiple authors, until Campbell and Lamar (2004) elevated *C. totonacus* to species status. Wüster et al., (2005) later showed that *C. totonacus* was in fact more closely related to members of the *Crotalus molossus* group than to *C. durissus*. Klauber (1952) revised the taxonomy of the *Crotalus durissus* group and described two subspecies based on coloration and morphology: *C. d. culminatus* (occurring in Michoacán, Morelos, Guerrero, and Oaxaca) and *C. d. tzabcan* (restricted to the Yucatan peninsula and Belize). Both subspecies were elevated to full species by Wüster et al. (2005) based on mtDNA data and morphological differences previously reported by Klauber (1952, 1972). Later, Campbell and Lamar (2004) split *Crotalus durissus* into two species: *Crotalus simus* Latreille 1801, which ranges from central Mexico to Costa Rica, and *C. durissus*, restricted to South America and a few continental islands. *Crotalus simus* contains

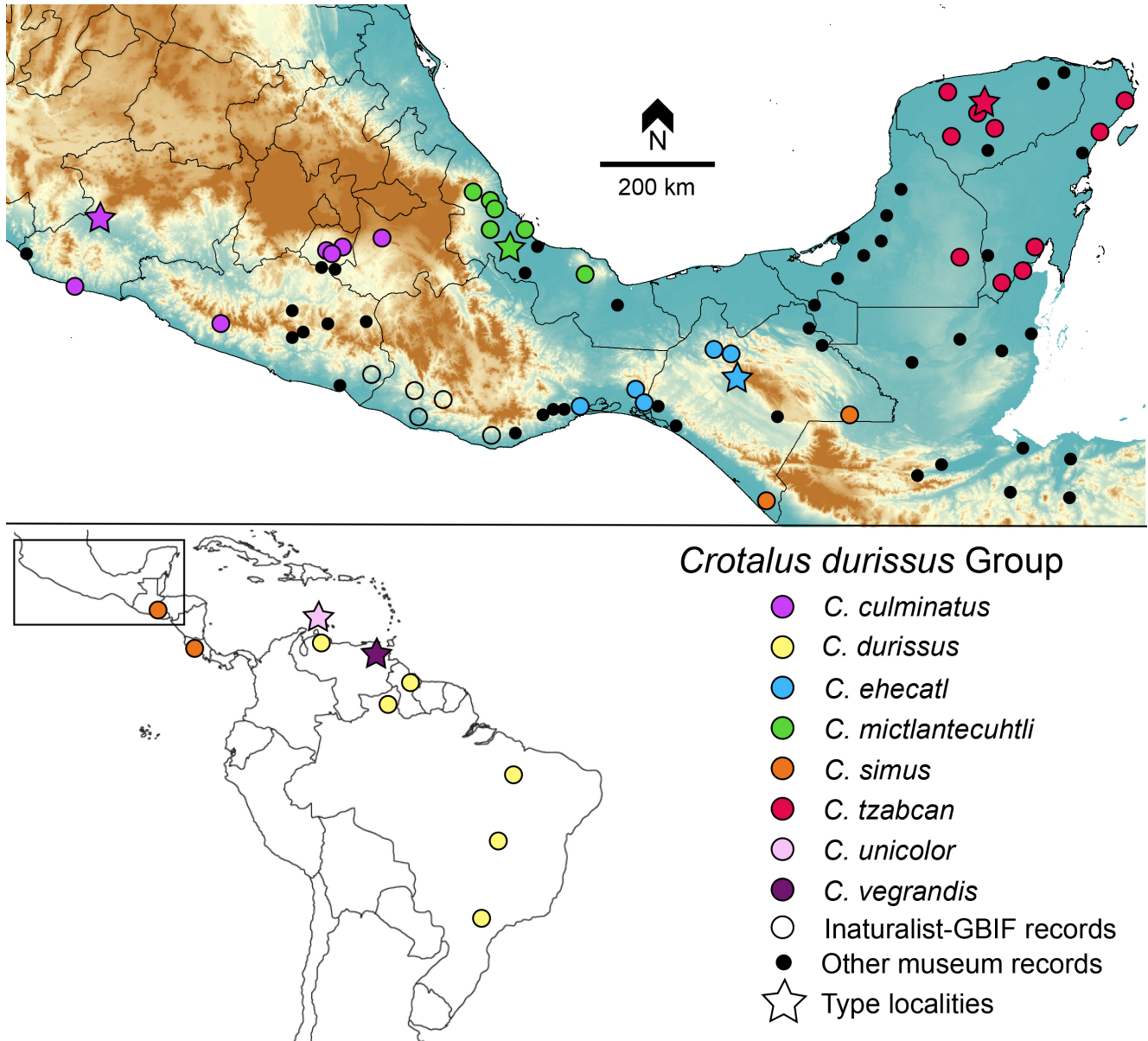


Figura 1. Mapa de distribución de los miembros del grupo de especies de *Crotalus durissus*. Las muestras con datos genéticos están representadas por círculos coloreados, mientras que otros registros sin datos genéticos disponibles se muestran como puntos negros. Los círculos abiertos indican registros adicionales para *C. culminatus* y *C. ehecattl* obtenidos de la base de datos gbif.org.

Figure 1. Distribution map of members of the *Crotalus durissus* species group. Samples with genetic data are represented by colored circles, while other museum records with no genetic data available are shown as black dots. Open circles indicate additional records for *C. culminatus* and *C. ehecattl* obtained from inaturalist.org as research grade observations, available in the gbif.org database. There is uncertainty regarding the type localities of *C. durissus* and *C. simus* and are thus not indicated.

no subspecies, whereas *C. durissus* contains nine subspecies: *C. durissus durissus* Linnaeus 1758, *C. d. cascavella* Wagler, 1824, *C. d. collilineatus* Amaral, 1926, *C. d. cumanensis* Humboldt, 1811, *C. d. marajoensis* Hoge, 1966, *C. d. maricelae* García-Perez, 1995, *C. d. ruruima* Hoge, 1966, *C. d. terrificus* (Laurenti, 1768) and *C. d. trigonicus* Harris & Simmons, 1978. Two other species from South

America *Crotalus unicolor* Van Lidth De Jeude, 1887 (restricted to Aruba) and *Crotalus vegrandis* Klauber, 1941 (restricted to Venezuela), have previously been considered subspecies of *C. durissus*, but are currently recognized by some taxonomist as full species (Wallach et al., 2014; Zaher et al., 2019).

Recently, Carbajal-Márquez et al. (2020) revised the taxonomy of the *C. durissus* group with additional sampling in Mexico. Based on mtDNA data and some morphological evidence, Carbajal-Márquez et al. (2020) split *C. culminatus* into three species, the nominal species *C. culminatus*, as well two new species, *Crotalus ehecatl* from southern Oaxaca and Chiapas, and *Crotalus mictlantecuhtli* from central Veracruz. These species were based on populations previously referred to as *C. simus culminatus* (*C. ehecatl*) and *C. simus simus* (*C. mictlantecuhtli*) by Campbell and Lamar (2004). Despite the additional sampling in the Carbajal-Márquez et al. (2020) study, their results do not necessarily show that some of these populations are indeed different species. Here we examine if the genetic patterns observed in previous studies could be explained by other biological processes, in particular, we test if isolation-by-distance (IDB) and a lack of thorough sampling might be the cause of an apparent barrier to geneflow between member of the *C. durissus* species group. Additionally, we test if a newly developed species delimitation method, DELINEATE (Sukumaran et al., 2021), supports the current taxonomy of the group.

MATERIALS AND METHODS

DNA sequencing and alignment. We sequenced the mitochondrial genes of Cytochrome B (Cytb) and NADH dehydrogenase subunit 4 (ND4) for five individuals of the *Crotalus durissus* group. These include one *C. culminatus*, one *C. ehecatl* and three *C. mictlantecuhtli* (Appendix 1). DNA extraction protocols and PCR conditions follow those of Reyes-Velasco et al., (2013) and were carried out at the Universidad Juárez del Estado de Durango. Additionally, we used the program MITObim (Hahn et al., 2013) to obtain mitochondrial sequences from the raw Illumina transcriptomic reads of Durban et al. (2017), which include two additional *C. mictlantecuhtli*, two *C. tzabcan*, and one *C. culminatus* (Appendix 1).

The program MITObim can generate mitochondrial contigs from short reads with the use of an iterative baiting method. We first used a published sequence of the mitochondrial genome of *Crotalus horridus* (GenBank reference number NC 014400) as the reference mitogenome, with the default program settings, except for a k-mer length of 31. We then repeated the same procedure for each individual, but with the resulting contig of the first run as the reference mitogenome. For the resulting mitogenomes, we extracted the Cytochrome B and ND4 genes for all subsequent analyses. All new mitochondrial sequences were deposited into GenBank (MZ855472-MZ855493; Appendix 1) and the following analyses are explained below.

As a backbone of our analyses, we used the sequences of Carbajal-Márquez et al. (2020). We obtained additional sequences from GenBank for the mitochondrial genes Cytb and ND4, as well as the nuclear gene oocyte maturation factor (*C-mos*) for most members in the *Crotalus durissus* and *C. molossus* species group. We reduced our dataset as to include only a single individual for most of the South American subspecies of *C. durissus*, as well as *C. unicolor* and *C. vegrandis*, since the main goal of this study is the northern members of the group. In the case of Mesoamerican specimens related to *C. simus*, we included all available individuals. Additionally, we included multiple other species of rattlesnakes as outgroups. A list of all sequences used and their GenBank number is presented in Appendix 1, while the locality for each sample is shown in figure 1. We aligned each individual gene using the online version of Mafft (Katoh & Standley, 2013). We then translated all genes to their predicted amino acid sequences in Geneious v9.1.6 (Biomatters Ltd., Auckland, NZ) to check for the presence of stop codons, which were not found.

Phylogenetic analyses. We estimated the best-fit models of nucleotide evolution for each gene and gene partition the program PartitionFinder. We then concatenated the individual genes using Sequence Matrix (Vaidya et al., 2011). We produced three datasets: one that included all genes (two mtDNA genes plus one nuclear gene), one that only included mtDNA, and the last one that only included the nuclear loci *c-mos*. We then used the Akaike Information Criterion (AIC) in the program PartitionFinder (Lanfear et al., 2012) to estimate partition schemes and models of molecular evolution, which are shown in table 1.

We estimated Bayesian phylogenetic inference (BI) for each dataset in Mr. Bayes v3.2. (Ronquist et al., 2012), in the CIPRES science gateway server (Miller et al., 2011). We used BI to estimate the posterior probabilities of phylogenetic trees based on a total of 10 million generations Metropolis-coupled Markov chain Monte Carlo (MCMC). These analyses consisted of four simultaneous runs, each with four chains (three heated and one cold), sampling every 1,000 generations. We verified that independent runs had converged by visualizing the output from BI in the program TRACER v. 1.5 (Drummond & Rambaut, 2007). Potential scale reduction factor (PSRF) estimates comparing chain likelihood values indicated convergence by one million generations, so we discarded the first 25% of BI samples as burn-in. We estimated a majority-rule consensus phylogram from the combination of the post burn-in samples from the four BI runs, combined the runs using TreeAnnotator v. 1.7.4 (Drummond & Rambaut, 2007) and visualized the resulting trees in FigTree



Tabla 1. Modelos de sustitución para los genes utilizados en este estudio.**Table 1.** Best-fits models of substitution for the genes used in this study.

Gene	Codon Position	Substitution Model
C-mos	1st	JC
C-mos	2nd	HKY
C-mos	3rd	HKY
CytB	1st	JC + gamma
CytB	2nd	JC + gamma
CytB	3rd	GTR + gamma
ND4	1st	JC + gamma
ND4	2nd	JC + gamma
ND4	3rd	GTR + gamma

v1.4.2 (Rambaut, 2014). Additionally, we used the NeighborNet algorithm in SplitsTree 4 (Huson & Bryant, 2006) to construct a phylogenetic network based on our alignment of mtDNA.

Genetic distances & Isolation-by-distance. In order to test for isolation-by-distance in multiple members of the *Crotalus durissus* group, first we georeferenced the locality data for all samples in our dataset with the use of Google Earth Pro. In a small number of cases, the locality information was incomplete or lacking and the locality information is tentative (for example, in the case of the single individual of *Crotalus unicolor*, the locality is “Aruba”, so we georeferenced this locality in the center of the island). Locality information and coordinates are provided in Appendix 1. We then obtained Euclidian distances between samples with the use of the earth.dist function in the R package fossil (Vavrek, 2011). We used the program MEGA X (Kumar et al., 2018) to estimate pairwise genetic distances (uncorrected P distances) of the mitochondrial data (Cytb + ND4), including all codon positions, both transitions and transversions, and Gamma distributed rates among sites. Genetic distances and geographic distances between samples are provided in Appendix 2 and 3. We assessed if isolation-by-distance (IBD) was present in the *C. durissus* group by testing for correlation between geographic distances and genetic distances, with the use of Mantel tests in the R package Vegan (Oksanen et al., 2013), as well as Multiple Matrix Regression with Randomization analysis, performed with the R function MMRR (Wang, 2013).

Additionally, we estimated pairwise genetic distances in ND4 for additional species of *Crotalus* which we obtained from

GenBank. We only included ND4 because this has traditionally been the most common gene used in rattlesnake systematics and was the only gene with a more thorough taxonomic coverage of the group. A list of all taxa and samples included is provided in Appendix 4.

Species delimitation analysis. To test previous species delimitations in the *Crotalus durissus* group, we employ a novel method DELINEATE (Sukumaran et al., 2021), which used a protracted speciation model (PSM). This method allows us to incorporate species limits obtain by other sources of data into the species delimitation analysis. However, this approach requires a good understanding of the current taxonomy of a particular group. The program's output is a topology in which each individual is assigned to either previously described species or to potentially new taxa.

First, we used BEAUTI, part of the BEAST software package (Drummond & Rambaut, 2007) to set up our StartBeast2 configuration file. For this analysis we used only the mitochondrial genes Cytb and ND4, with a single strict clock model of substitution across genes, and a HKY+G substitution model. Due to the computational burden of the analysis, we reduced our dataset to include only four individuals per species and additionally included multiple individuals of the *Crotalus molossus* species group, which are sister to the *C. durissus* group (Reyes-Velasco et al., 2013). We ran four replicates of the analysis, which consisted of 400 million generations, sampling every 400,000 generations, for a total of 1000 trees.

We then used the sumtrees.py script, part of the DendroPy library (Sukumaran & Holder, 2010) to discard the first 250 trees as burn-in, combine the runs and use the Maximum Clade Credibility Tree to summarize the resulting trees. We then prepared our population assignment input file for DELINEATE and constrained all currently recognized species, except for *Crotalus ehecatl*, *C. mictlantecuhtli*, *C. simus*, *C. unicolor* and *C. vegrandis*. We tested these last two species, as preliminary phylogenetic analysis showed that they were nested within *C. durissus* (see below), so we wanted to test their validity as species. In the case of the *Crotalus molossus* group, we constrained all species, except for the three subspecies of *C. molossus* (*C. m. molossus*, *C. m. nigrescens* & *C. m. oaxacus*) to test their species assignment.

RESULTS

Alignments. Our resulting alignments for each gene were the following: 751bp (Cytb), 663bp (ND4) and 594bp (c-mos). In the

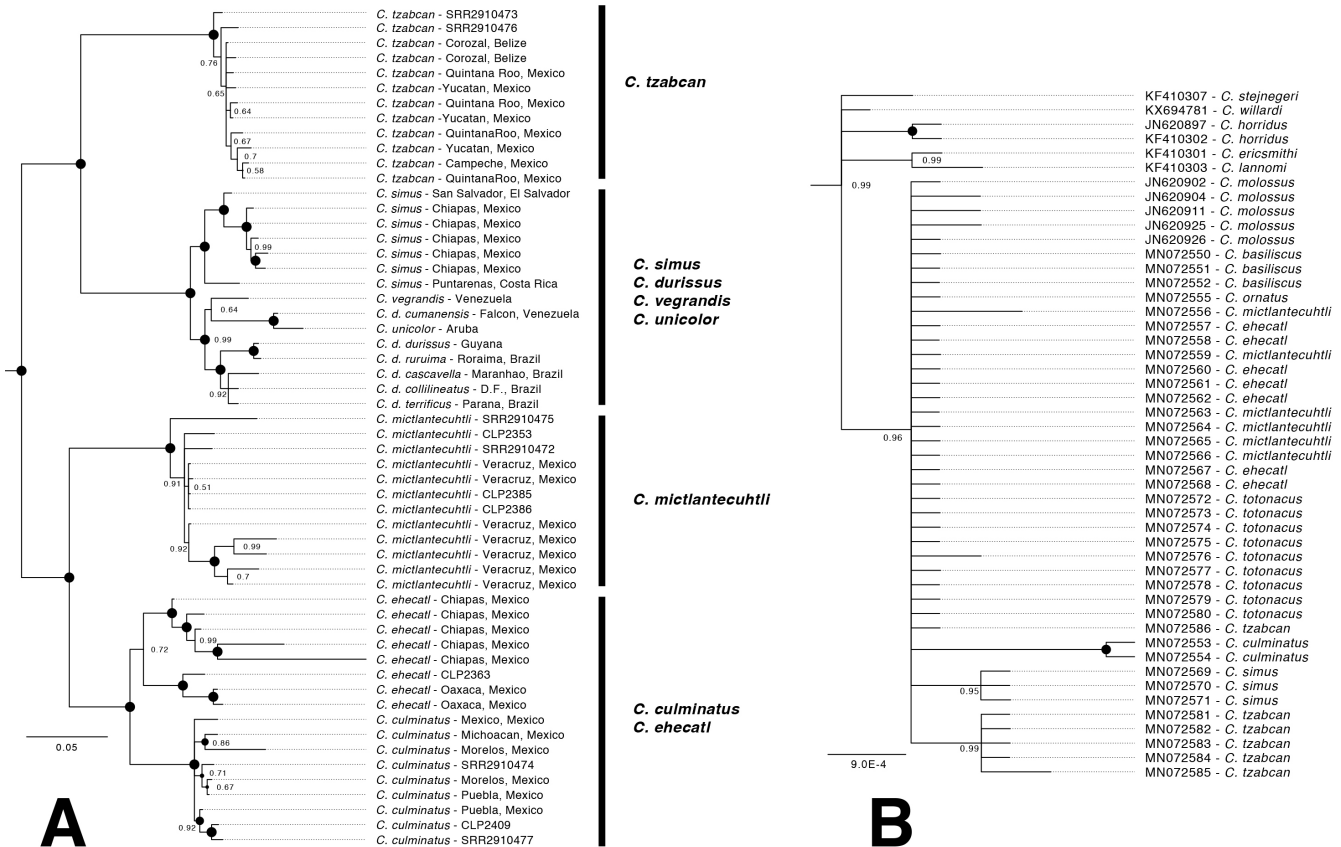


Figura 2. Inferencia filogenética de los miembros del grupo de especies de *Crotalus durissus*. A) Inferencia filogenética bayesiana basada en los genes de ADN mitocondrial concatenados Cytb y ND4. B) Inferencia filogenética bayesiana basada en los locus nuclear C-mos. Los círculos negros representan nodos con un soporte posterior de 1.

Figure 2. Phylogenetic inference of members of the *Crotalus durissus* species group. A) Bayesian phylogenetic inference based on the concatenated mtDNA genes Cytb and ND4. B) Bayesian phylogenetic inference based on the nuclear locus C-mos. Black circles represent nodes with a posterior support of 1.

case of the mtDNA genes, the number of parsimony-informative sites was 107 for Cytb and 162 for ND4, which represents 14% and 24% of all sites, respectively. In the case of c-mos, only 19 sites were parsimony-informative, which represents only 3% of all sites.

Phylogenetic analyses. Our phylogenetic analyses of the mtDNA and mtDNA + c-mos datasets resulted in identical topologies, which are in concordance with other studies of the same group, which is not surprising as the sequences used in the different studies are nearly the same, and any signal in the nuclear gene c-mos becomes overwritten by the mtDNA data, thus here we only present the mtDNA phylogeny (Fig. 2A). We recovered four main clades in the *C. durissus* group: The first clade included all individuals of *Crotalus tzabcan* from the Yucatan peninsula and Belize and received strong support (posterior probability, pp = 1). This clade was sister to a second well supported clade which

consisted of *Crotalus durissus* (all subspecies), *C. vegrandis* and *C. unicolor* (which were nested within *C. durissus*), as well as all *C. simus* from southern Chiapas and Central America. The third clade consisted of individuals of *C. mictlantecuhtli* from Veracruz and was sister to the last clade which was composed of *C. culminatus* plus *C. ehecatl*. When we analyzed the nuclear gene c-mos alone, we recover a polytomy which included most members of the *C. durissus* group, as well as other species of rattlesnakes (Fig 2B). These results show that c-mos lacks phylogenetic signal and thus is of little use for solving the phylogenetic relationships of the *C. durissus* group.

mtDNA gene network & genetic distances. When we visualized the mtDNA data as a genetic network in SplitsTree (Fig 3), we recovered the same general patterns as in the Bayesian Inference analysis of mtDNA, however the degree of low genetic variation between many of populations in the *Crotalus durissus* group were

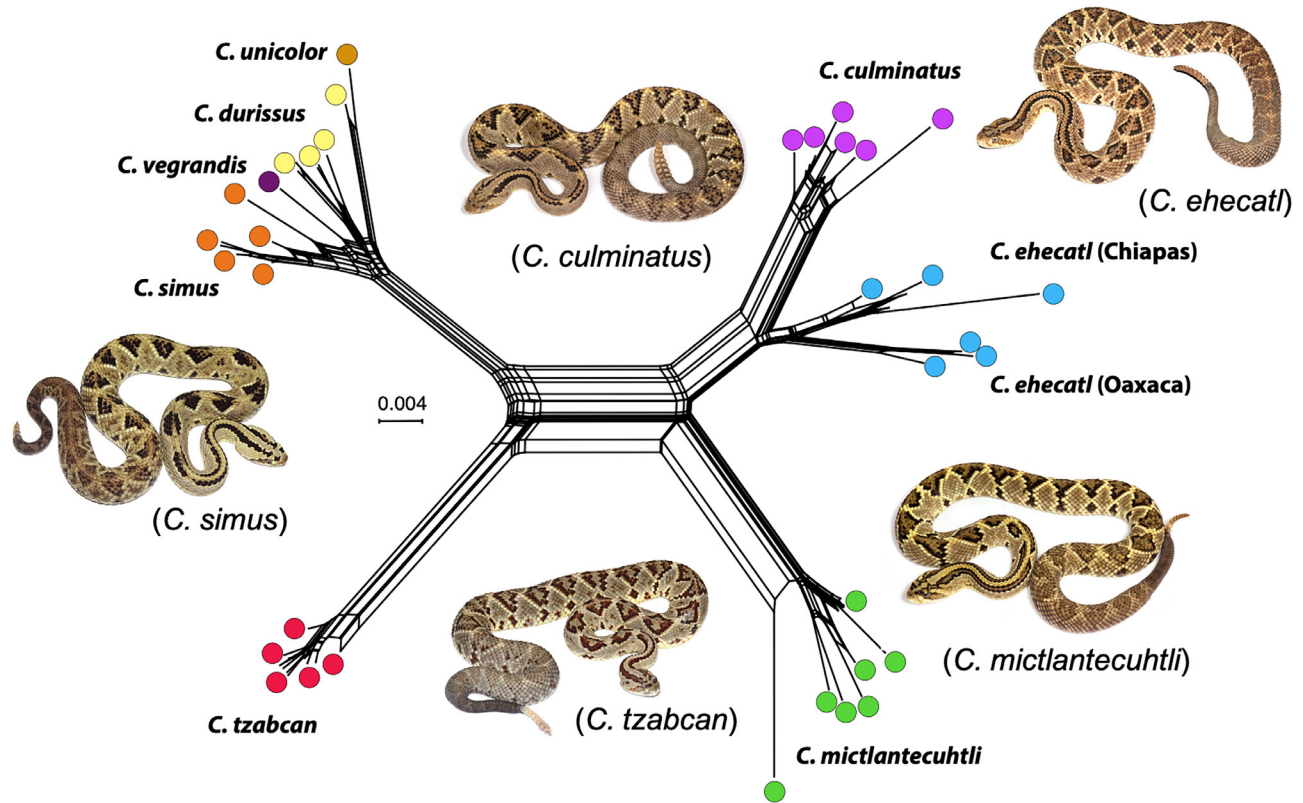


Figura 3. Splits tree network de los genes de los genes mitocondriales concatenados Cytb y ND4 de los miembros del grupo de especies de *Crotalus durissus*. Las imágenes representan miembros del grupo de especies de *C. durissus*.

Figure 3. Splits tree network of the concatenated mtDNA genes Cytb and ND4 of members of the *Crotalus durissus* species group. Images represent multiple species of the *C. durissus* group.

more obvious (at least at the mtDNA level). All the subspecies of *C. durissus*, as well as *C. unicolor*, *C. vegrandis* and *C. simus* formed a single cluster, while both *C. tzabcan* and *C. mictlantecuhtli* formed independent clusters. In the case of *C. culminatus* and *C. ehecatl*, they formed a single cluster with some subdivisions, with individuals of *C. culminatus*, *C. ehecatl* from Chiapas and *C. ehecatl* from Oaxaca forming subclusters. These subcluster could be the result of Isolation-by-distance (see below).

All the subspecies of *Crotalus durissus*, including *C. unicolor* and *C. vegrandis* had genetic distances (uncorrected p distances) of only 0.018 - 0.036 when compared to populations of *C. simus* from Mexico and Central America (Table 1) Similarly, *C. culminatus* and *C. ehecatl* had very limited genetic differences between each other, ranging from 0.026 to 0.049. These genetic distances are similar to those shown by intraspecific populations of other rattlesnakes (Appendix 4). On the other hand, *C. tzabcan*, which is not clearly isolated from other members of the group in northern Central America or central Veracruz, showed

rather significant genetic differences with all other members of the group, ranging from 0.06 to 0.9, which are on par with interspecific differences in other rattlesnake taxa (Appendix 4).

Isolation-by-distance. Both Mantel tests and MMRR analyses showed a positive correlation between genetic distances and geographic distances in six out of 10 tests we performed ($p=0.001$; Fig. 4 & Appendix 5). The Mantel test showed a strong positive correlation between genetic and geographic distances (Mantel statistic (r) >0.5) when comparing the following species: *Crotalus ehecatl* - *C. mictlantecuhtli* ($r=0.82$), *C. culminatus* + *C. ehecatl* - *C. tzabcan* ($r=0.72$), *Crotalus culminatus* - *C. mictlantecuhtli* ($r=0.66$) and *C. culminatus* - *C. ehecatl* ($r=0.62$). However, in the MMRR analysis, the correlation was still significant, but not as strong in those same species pairs. When *C. culminatus*, *C. ehecatl* and *C. mictlantecuhtli* were included, the relationship between genetic distance and geographic distance was significant, but not strong (Mantel statistic (r) = 0.44; MMRR (r) = 0.1966).

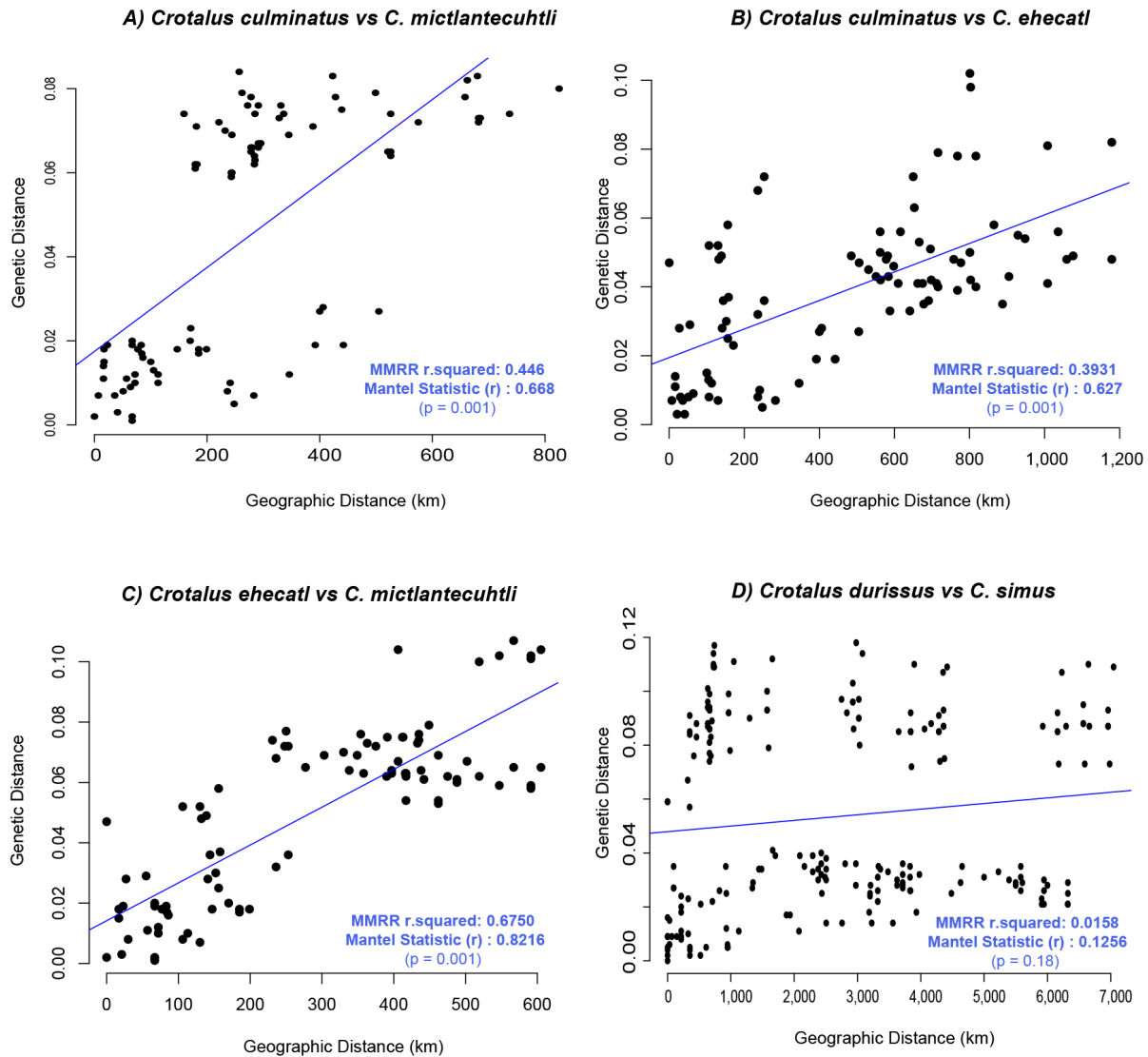


Figura 4. Gráfica de aislamiento por distancia (APD) que muestra la relación entre las distancias geográficas y genéticas de los miembros del grupo de especies de *Crotalus durissus*. La línea representa la correlación entre la distancia geográfica y la distancia genética (r). La significancia estadística se estimó con una prueba de Mantel.

Figure 4. Isolation by distance (IBD) plot showing relationship between geographic and genetic distances for members of the *Crotalus durissus* species group. Line represents correlation between geographic and genetic distance (r). Statistical significance was estimated with a Mantel test.

If only individuals of *C. culminatus* were analyzed, the relationship was strong in the Mantel test (Mantel statistic (r) = 0.66), but not in the MMRR analysis (r = 0.4355), however, neither the Mantel test or the MMRR analysis was significant (p = 0.058; p = 0.057, respectively). When comparing *Crotalus simus* and *C. durissus* (including *C. vegrandis* and *C. unicolor*), the relationship was weak or non-existent (Mantel statistic (r) = 0.12; MMRR (r) = 0.01) and not significant (p = 0.132; p = 0.18, respectively). Similar weak and non-significant relationships were found when including only individuals of *C. tzabcan* (Mantel

statistic (r) = 0.03; MMRR (r) = 0.0009) or *C. durissus* (including *C. vegrandis* and *C. unicolor*; Mantel statistic (r) = 0.246; MMRR (r) = 0.6). These results show that at least in some member of the *C. durissus* group, isolation-by-distance might be responsible of the genetic structure observed in the mitochondrial genes.

Species delimitation analysis. The results of our species delimitation analysis in DELINEATE supported the recognition of *Crotalus mictlantecuhtli* as a distinct species, on the other hand, it included the samples of *C. ehecatl* as part of *C. culminatus*

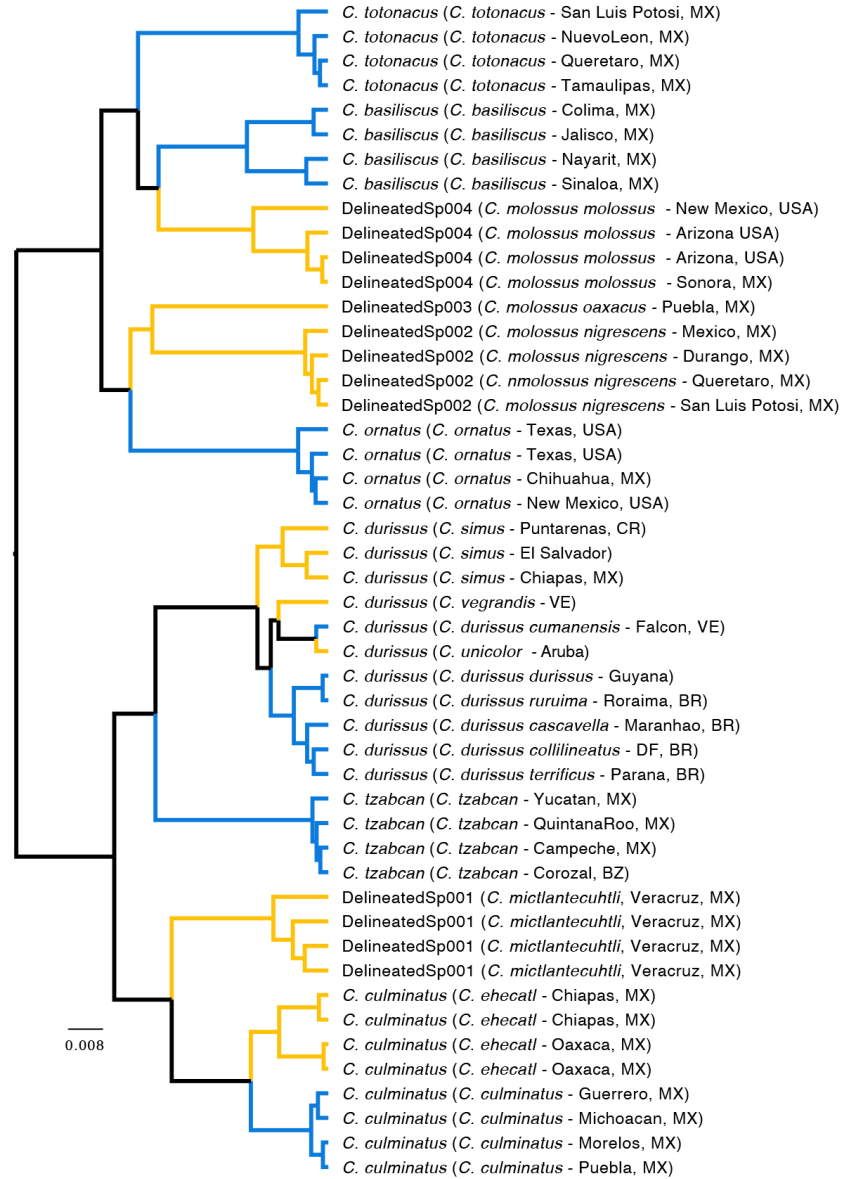


Figura 5. Resultados del análisis de delimitación de especies realizado en DELINEATE. Los clados en azul representan las asignaciones a especies, mientras que los clados en amarillo representan a los linajes cuya asignación a alguna especie fue examinada. Los nombres en las puntas del árbol que aparecen como "DelineatedSp" representan especies que fueron probadas y que el programa reconoce como distintas, mientras que los nombres entre paréntesis representan su anterior asignación a especies y localidad.

Figure 5. Results of the species delimitation analysis performed in DELINEATE. Clades in blue represent constrained species, while clades in yellow represent those lineages whose species designations were tested. Tips with species assignment as "DelineatedSp" represent species that were tested which the program recognizes as distinct, while names in parenthesis represent their previous species assignment and locality.

(Fig. 5). Additionally, this analysis recovered all individuals of *C. simus*, *C. unicolor* and *C. vegrandis* as conspecific with *C. durissus*. In the case of the *C. molossus* group (which we included as a comparison), we recovered each of the tree subspecies of *C. molossus* (*C. m. molossus*, *C. m. nigrescens* & *C. m. oaxacus*) as full species.

DISCUSSION

In this study we evaluate if isolation-by-distance could be responsible for the genetic variation observed at the mitochondrial level in members of the *Crotalus durissus* species group. We do this by sequencing additional individuals,

combined with previously published mitochondrial and nuclear data. Our results show that the genetic differences between some of the species in the group (*C. simus* – *C. durissus* and *C. culminatus* – *C. ehecatl*) are lower than intra-specific genetic distances (Appendix 2). The only nuclear gene at hand for the *C. durissus* group (*c-mos*) is not informative for distinguishing between many species of rattlesnakes (Fig. 2B), and thus it is of little use when it comes to taxonomic decisions. Additionally, in the case of *Crotalus ehecatl* and *C. culminatus*, morphological differences were also minimal, and the characters analyzed overlapped almost completely (see Carbajal-Márquez et al., 2020, Fig. 4).

Isolation-by-distance (IBD) is often overlooked in population genetics (Meirmans, 2012). In the case of some members of the *Crotalus durissus* group, IBD might be responsible for the apparent genetic differences between populations, which have previously been inferred to be inter-specific differences (Fig. 4). The individuals of *C. culminatus* and *C. ehecatl* from which mtDNA sequences were analyzed are separated by more than 950 km (coast of Michoacán vs Oaxaca-Chiapas border; Fig. 1). There are records for *C. culminatus* along the Pacific coast of Mexico, from the Colima-Michoacán border to eastern Oaxaca (Campbell & Lamar, 2004), followed by an apparent gap of approximately 300 km between Marquelia, Guerrero and Santa Maria Huatulco, Oaxaca (the western most record *C. ehecatl*). However, multiple research grade observations for *Crotalus* exist along the coast of Oaxaca in the Inaturalist database (Fig. 1), which shows that the distribution of *C. culminatus* is continuous and no gap exist between *C. culminatus* and *C. ehecatl*. Unfortunately, no DNA sequences exist for these samples. Our analyses of IBD show that there is a strong correlation between genetic and geographic distances in the *C. culminatus* group (Fig. 4 & Appendix 5), and thus could be the cause of the apparent genetic distinctiveness between *C. culminatus* and *C. ehecatl*. Despite the fact that we did not find evidence for IBD between *C. simus* and *C. durissus*, it is still possible that IBD exists, but it is not detectable with mitochondrial markers alone (Teske et al., 2018). Several recent studies have shown that if IBD is not taken into account, genetic differences can drastically affect the results of species delimitation methods, and over-split species (Gratton et al., 2016; Mason et al., 2020), which could be the case in some members of the *C. durissus* group.

We use the program DELINEATE as a way to test for current species hypothesis in the group. This analysis did not recognize *Crotalus ehecatl* as distinct from *C. culminatus* but did recover *C. mictlantecuhtli* as a separate lineage. DELINEATE also failed to recover *C. simus*, *C. unicolor* and *C. vegrandis* as distinct from

C. durissus. However, all these results should be taken with caution, as our analysis included only two mitochondrial genes, and hybridization or mitochondrial introgression could be responsible for this patterns. Campbell and Lamar (2004) split *Crotalus simus* and *C. durissus* into two different species. These species are separated by ~900 km in Central America, and both appear to be absent in Panamá. However, the mtDNA differences between these species are minimal, and much smaller than population level differences in other species (see Appendix 4). Wüster et al. (2005) and Quijada-Mascareñas et al., (2007) suggested that the invasion of South America by the *C. durissus* occurred relatively recently (~1 mya), probably facilitated by a continuous corridor of dry savannahs. However, the divergence time estimated by Wüster et al. (2005) for the split between the *C. durissus* and the *C. molossus* groups is much older (10-14 mya) than that estimated by Reyes-Velasco et al., (2013) of ~3-6 mya, however, these divergence dates should be taken with caution. A much younger split between these groups could indicate that the invasion of South America by *C. durissus* might be younger than expected, and probably occurred faster than previously suggested, which would explain the almost non-existent differences in the mtDNA data and the results of the species delimitation analysis.

CONCLUSION

Species are, by definition, hypotheses (De Queiroz, 2007; Samadi & Barberousse, 2006), therefore, the more evidence in favor of a hypothesis, the better species is supported. Just as when testing any hypothesis, careful consideration should be taken into account when describing new species. It is relatively easy to describe new taxa, especially when only a few molecular markers are analyzed for diagnosing between species. However, revising the evidence for such taxonomic changes can be very time consuming, and many people are not willing, nor have the time for these taxonomic revisions. Therefore, before describing new taxa, is important to consider all available evidence in favor or against any taxonomic designations. We do not argue that differences in mtDNA or on a few markers should be used as a yardstick to define what species are valid and what species are not, on the contrary, we argue against this type of approach, and thus we understand the limitations of our own analyses. Therefore, our goal with this study is not to conclude how many species should be considered valid in the *Crotalus durissus* species group, but to show that the data at hand can be interpreted in multiple ways. We believe that additional sampling, combined with more thoroughly molecular analyses are necessary to better understand gene flow and species limits in the *C. durissus* group, especially in areas of possible contact such as the southern part

of the Yucatan Peninsula, Chiapas, Guatemala, Tabasco and the Chimalapas region of Oaxaca and Chiapas (Fig. 1).

Acknowledgments.— We would like to thank Ruben Carbajal-Márquez, Jesus Sigala-Rodriguez, and Christoph I. Grünwald for interesting discussions that greatly improved the quality of this paper. Javier Ortiz-Medina kindly provided images of rattlesnakes. Comments from two anonymous reviewers greatly improved an earlier version of the manuscript.

SUPPLEMENTARY INFORMATION

This article contains supplementary material available at (<https://drive.google.com/drive/folders/1PwIQtsnzRD1bn4py4nJLGVZbQQYokl5?usp=sharing>)

Appendix I. Catalogue numbers, GenBank numbers and locality information for samples used in this study.

Apéndice I. Números de catálogo, números de Genbank e información de localidad de ejemplares utilizados en este estudio.

Appendix II. Genetic distances (uncorrected p distances) for members of the *Crotalus durissus* species group, estimated from the concatenated mtDNA genes Cytb and ND4.

Apéndice II. Distancias genéticas (distancias p no corregidas) para los miembros del grupo de especies de *Crotalus durissus*, estimadas a partir de los genes mitocondriales concatenados Cytb y ND4.

Appendix III. Geographic distances (Km) for members of the *Crotalus durissus* species group.

Apéndice III. Distancias geográfica (Km) para miembros del grupo de especies de *Crotalus durissus*.

Appendix IV. Genetic distances (uncorrected p distances) for multiple species and populations of the genus *Crotalus*, estimated from the mtDNA gene ND4.

Apéndice IV. Distancias genéticas (distancias p no corregidas) de múltiples especies y poblaciones del género *Crotalus*, estimadas del gen mitocondrial ND4.

Appendix V. Results of the Mantel test for correlation between genetic distance and geographic distance in members of the *Crotalus durissus* species group.

Apéndice V. Resultados de la prueba de Mantel para la correlación entre la distancia genética y la distancia geográfica en los miembros del grupo de especies de *Crotalus durissus*.

CITED LITERATURE

- Campbell, J.A. & W.W. Lamar. 2004. The Venomous Reptiles of the Western Hemisphere (Vol. 2). Comstock Pub. Associates, Ithaca, New York, USA.
- Carbajal-Márquez, R.A., J.R. Cedeño-Vázquez, A. Martínez-Arce, E. Neri-Castro & S.C. Machkour-M'Rabet. 2020. Accessing cryptic diversity in Neotropical rattlesnakes (Serpentes: Viperidae: *Crotalus*) with the description of two new species. *Zootaxa* 4729:451-481.
- Chambers, E.A. & D.M. Hillis. 2020. The multispecies coalescent over-splits species in the case of geographically widespread taxa. *Systematic Biology* 69:184-193.
- De Queiroz, K. 1998. The general lineage concept of species, species criteria, and the process of speciation. Pp. 57-75. En: D.J. Howard & S.H. Berlocher (Eds.). *Endless Forms: species and speciation*. Oxford University Press, New York, USA.
- De Queiroz, K. 2007. Species concepts and species delimitation. *Systematic Biology* 56:879-886.
- Drummond, A.J. & A. Rambaut. 2007. BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology* 7:1-8.
- Durban, J., L. Sanz, D. Trevisan-Silva, E. Neri-Castro, A. Alagon & J.J. Calvete. 2017. Integrated venomomics and venom gland transcriptome analysis of juvenile and adult mexican rattlesnakes *Crotalus simus*, *C. tzabcan*, and *C. culminatus* revealed miRNA-modulated ontogenetic shifts. *Journal of Proteome Research* 16:3370-3390.
- Fujita, M.K., A.D. Leaché, F.T. Burbrink, J.A. McGuire & C. Moritz. 2012. Coalescent-based species delimitation in an integrative taxonomy. *Trends in Ecology & Evolution* 27:480-488.
- Gloyd, H.K. & C.F. Kauffeld. 1940. A new rattlesnake from Mexico. *Chicago Academy of Sciences* 6:11-14.
- Hahn, C., L. Bachmann & B. Chevreaux. 2013. Reconstructing mitochondrial genomes directly from genomic next-generation sequencing reads—a baiting and iterative mapping approach. *Nucleic Acids Research* 41(13):e129.



- Hillis, D.M. 2019. Species delimitation in herpetology. *Journal of Herpetology* 53:3-12.
- Huson, D.H. & D. Bryant. 2006. Application of phylogenetic networks in evolutionary studies. *Molecular Biology and Evolution* 23:254-267.
- Katoh, K. & D.M. Standley. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution* 30, 772-780.
- Klauber, L.M. 1952. Taxonomic studies on rattlesnakes of Mainland Mexico. *Bulletins of the Zoological Society of San Diego* 26:1-143.
- Klauber, L.M. 1972. *Rattlesnakes: Their Life History, Habits, and Influence on Mankind*. University of California Press, Berkeley, USA.
- Kumar, S., G. Stecher, M. Li, C. Knyaz & K. Tamura 2018. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution* 35(6):1547-1549.
- Lanfear, R., B. Calcott, S.Y. Ho & S. Guindon. 2012. PartitionFinder: combined selection of partitioning schemes and substitution models for phylogenetic analyses. *Molecular Biology and Evolution* 29:1695-1701.
- Meirmans, P. G. 2012. The trouble with isolation by distance. *Molecular Ecology* 21:2839-2846.
- Miller, M.A., W. Pfeiffer & T. Schwartz. 2011. The CIPRES science gateway: a community resource for phylogenetic analyses. Pp. 1-8. En: *Proceedings of the 2011 TeraGrid Conference: Extreme Digital Discovery*.
- Oksanen, J., F.G. Blanchet, R. Kindt, P. Legendre, P.R. Minchin, R. O'hara, G.L. Simpson, P. Solymos, M.H.H. Stevens, H. Wagner & M.J. Oksanen 2013. Package 'vegan'. *Community ecology package, version 2.1-295*.
- Padial, J.M., A. Miralles, I. De la Riva & M. Vences. 2010. The integrative future of taxonomy. *Frontiers in Zoology* 7:1-14.
- Pante, E., C. Schoelinc & N. Puillandre. 2015. From integrative taxonomy to species description: one step beyond. *Systematic Biology* 64:152-160.
- Quijada-Mascareñas, J.A., J.E. Ferguson, C.E. Pook, M.D.G. Salomao, R.S. Thorpe & W. Wüster. 2007. Phylogeographic patterns of trans-Amazonian vicariants and Amazonian biogeography: the Neotropical rattlesnake (*Crotalus durissus* complex) as an example. *Journal of Biogeography* 34:1296-1312.
- Rambaut, A. 2014. FigTree v1. 4.2. <http://tree.bio.ed.ac.uk/software/figtree/>. [Accessed on September 2021].
- Reyes-Velasco, J., J.M. Meik, E.N. Smith & T.A. Castoe 2013. Phylogenetic relationships of the enigmatic longtailed rattlesnakes (*Crotalus ericsmithi*, *C. lannomi*, and *C. stejnegeri*). *Molecular Phylogenetics and Evolution* 69:524-534.
- Ronquist, F., M. Teslenko, P. Van Der Mark, D.L. Ayres, A. Darling, S. Höhna, S., B. Larget, L. Liu, M.A. Suchard & J.P. Huelsenbeck. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61:539-542.
- Rubioff, D. & B.S. Holland. 2005. Between two extremes: mitochondrial DNA is neither the panacea nor the nemesis of phylogenetic and taxonomic inference. *Systematic Biology* 54:952-961.
- Samadi, S. & A. Barberousse. 2006. The tree, the network, and the species. *Biological Journal of the Linnean Society* 89:509-521.
- Smith, H.M. & E.H. Taylor. 1945. An annotated checklist and key to the snakes of Mexico. *Bulletin of the United States National Museum* 187:1-239.
- Sukumaran, J. & M.T. Holder. 2010. DendroPy: a Python library for phylogenetic computing. *Bioinformatics* 26:1569-1571.
- Sukumaran, J., M.T. Holder & L.L. Knowles. 2021. Incorporating the speciation process into species delimitation. *PLoS Computational Biology* 17:e1008924.
- Sukumaran, J. & L.L. Knowles. 2017. Multispecies coalescent delimits structure, not species. *Proceedings of the National Academy of Sciences* 114:1607-1612.
- Teske, P.R., T.R. Golla, J. Sandoval-Castillo, A. Emami-Khoyi, C.D. Van der Lingen, S. Von der Heyden, B. Chiazzari, B.J. Van Vuuren, & L.B. Beheregaray. 2018. Mitochondrial DNA is unsuitable to test for isolation by distance. *Scientific Reports* 8:1-9.
- Ueda K. 2021. iNaturalist Research-grade Observations. iNaturalist.org. Occurrence dataset <https://doi.org/10.15468/ab385x> [Accessed on October, 2021].



- Vaidya, G., D.J. Lohman & R. Meier. 2011. SequenceMatrix: concatenation software for the fast assembly of multi-gene datasets with character set and codon information. *Cladistics* 27:171-180.
- Vavrek, M.J. 2011. Fossil: palaeoecological and palaeogeographical analysis tools. *Palaeontologia Electronica* 14:16.
- Wallach, V., K.L. Williams & J. Boundy. 2014. *Snakes of the world: a catalogue of living and extinct species*. CRC press, New York, USA.
- Wang, I.J. 2013. Examining the full effects of landscape heterogeneity on spatial genetic variation: a multiple matrix regression approach for quantifying geographic and ecological isolation. *Evolution* 67:3403-3411.
- Will, K.W., B.D. Mishler & Q.D. Wheeler. 2005. The perils of DNA barcoding and the need for integrative taxonomy. *Systematic Biology* 54:844-851.
- Wüster, W., J.E. Ferguson, J.A. Quijada-Mascareñas, C.E. Pook, M. Da Graca Salomao & R.S. Thorpe. 2005. Tracing an invasion: landbridges, refugia, and the phylogeography of the Neotropical rattlesnake (Serpentes: Viperidae: *Crotalus durissus*). *Molecular Ecology* 14:1095-1108.
- Zaher, H., R.W. Murphy, J.C. Arredondo, R. Graboski, P.R. Machado-Filho, K. Mahlow, G.G. Montingelli, A.B. Quadros, N.L. Orlov, M. Wilkinson, Y.P. Zhang & F.G. Grazziotin. 2019. Large-scale molecular phylogeny, morphology, divergence-time estimation, and the fossil record of advanced caenophidian snakes (Squamata: Serpentes). *PloS One* 14:e0216148.

